Development of Tools by EPA to Determine the Effectiveness of Green Infrastructure-Based Approaches to Mitigate Stormwater

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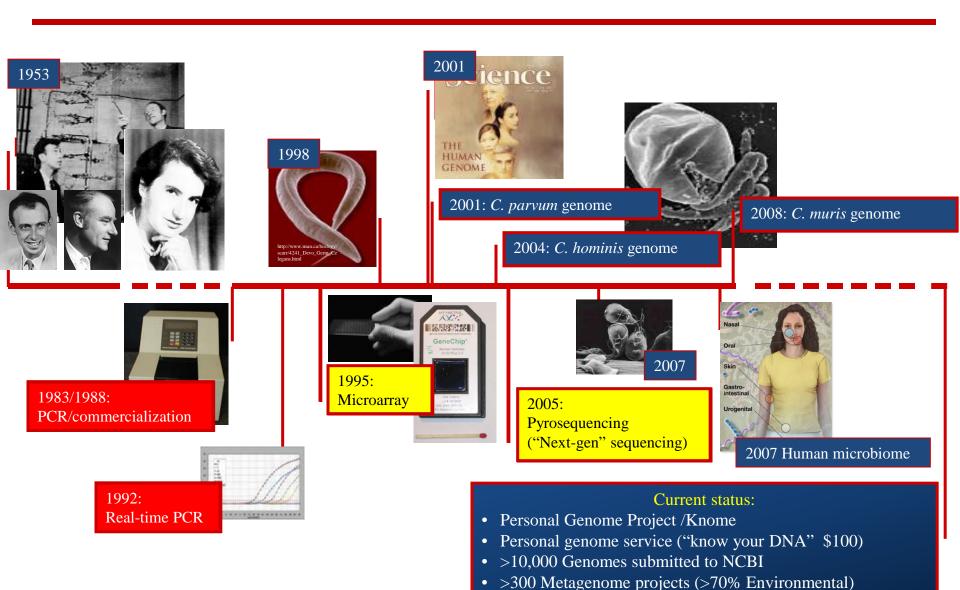
Acknowledgement of the team:

- Nick Ashbolt
- Ann Grimm
- Kevin Oshima
- Nichole Brinkman
- Scott Keely
- Shay Fout
- Rich Haugland
- Eric Villegas
- Brian Zimmerman
- Daniel Divelbiss
- All from the Microbiological and Chemical Exposure Assessment Research Division (MCEARD)

Overview of Talk

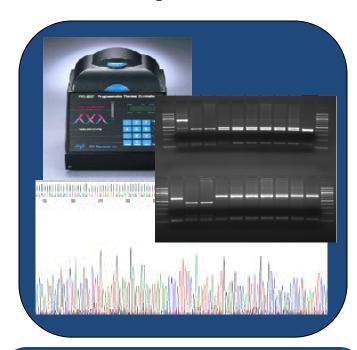
- Molecular Context of Tool Development
- General Approach for assessing microbial risks associated with water reuse
 - Performance assessment of treatment via spiking
 - Validate indicators by comparison with pathogens in mixed spiked cocktails
 - Goals
 - Controlled testing to define best management practices
 - Potentially develop real time or near real time monitoring (perhaps)
 - Where appropriate (source, use effects)
 - Pathogens are a difficult target
 - Biologically-based indicators are less difficult but still emerging
 - On-line process performance measures linked to BMP definition
 - Future of green infrastructure

$DNA \rightarrow PCR \rightarrow Genomes$

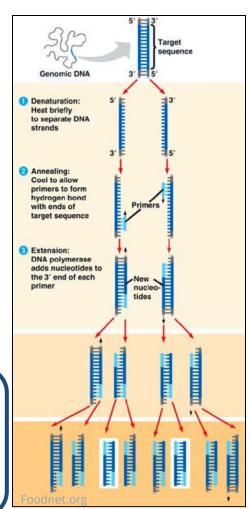


End-point vs. real-time PCR

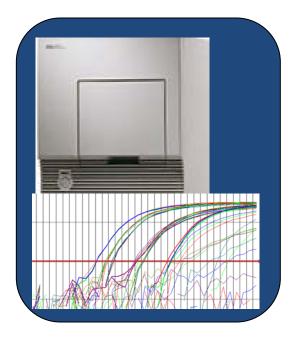
End-point PCR



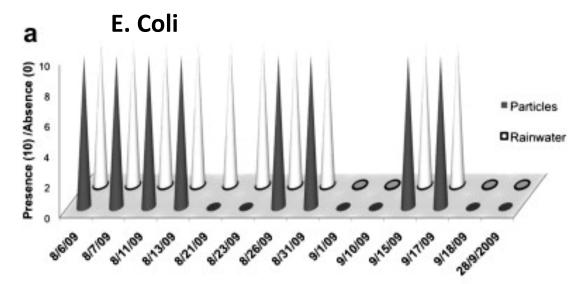
- Semi-quantitative (densitometry)
- Can amplify longer sequences
- Very specific
- Sequencing compatible

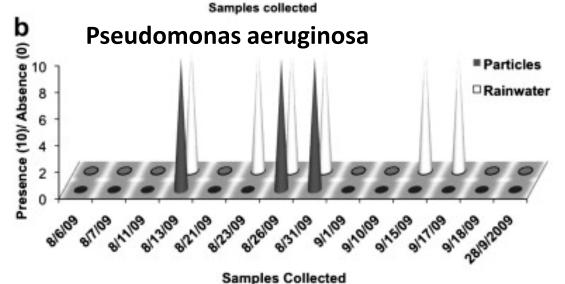


Real-time PCR



- Quantitative/standard curve
- Fluorescent probe
- Short PCR product (amplicon)
- Very specific





Assessment of bacterial pathogens in fresh rainwater and airborne particulate matter using Real-Time PCR

New tools expand sampling possibilities

Data still "noisy" event driven

Kaushik & Balasubramanian
Atmospheric Environment 46:131

Correlations between fecal indicators and pathogens in rainwater tanks in Australia

P value for correlation to the indicated pathogen

Poor correlation

reca indicat bacteria	New Indicators and new approach needed!						
	А.			L.		G. lamblia	
	<i>hydrophila</i> <i>lip</i> gene	C. coli ceuE gene	<i>C. jejuni</i> mapA gene	<i>pneumophil</i> <i>a mip</i> gene	Salmonella invA gene	β-giardin gene	
E. coli	0.250	0.611	0.466	0.969	0.306	0.406	
Enterococci	0.020 <u>b</u>	0.142	0.552	0.878	0.986	0.873	
C. perfringens	0.759	0.752	0.909	0.469	0.107	0.316	

Ahmed et al. 2008 AEM 74:5490

STEP 1
SETTING

Problem formulation & Hazard identification

Describe physical system, selection of reference pathogens and identification of hazardous events

Quantitative microbial risk STEP 2 risk exposure assessment (QMRA)

Rain / Storm water
Pathogen concentrations

Ingress
Ingress pathogen

(Pingress)

Cistern storage
Pathogen loss
(sediment/biofilm/death)

Treatment (UV/CI₂)
Pathogen removal

Non-Potable exposures
Volume water consumed

STEP 3
HEALTH EFFECTS

STEP 4

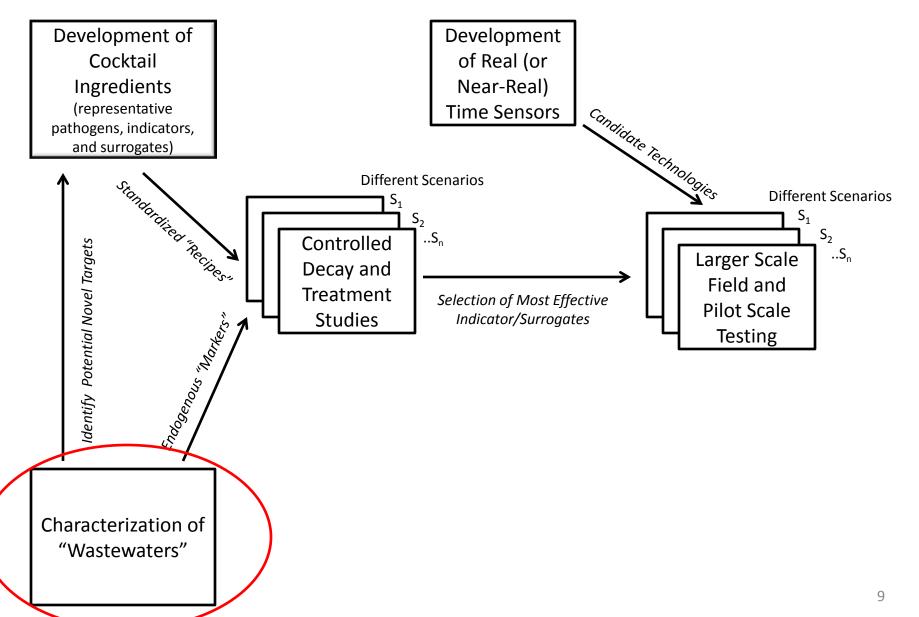
Dose-Response (Pinf)

Selection of appropriate models for each pathogen and the population exposed

Risk Characterisation

Simulations for each pathogen baseline and event infection risks with variability & uncertainty identified

Overall Research Plan for Developing Tools for Assessing Efficacy of Water Reuse Approaches



Summary

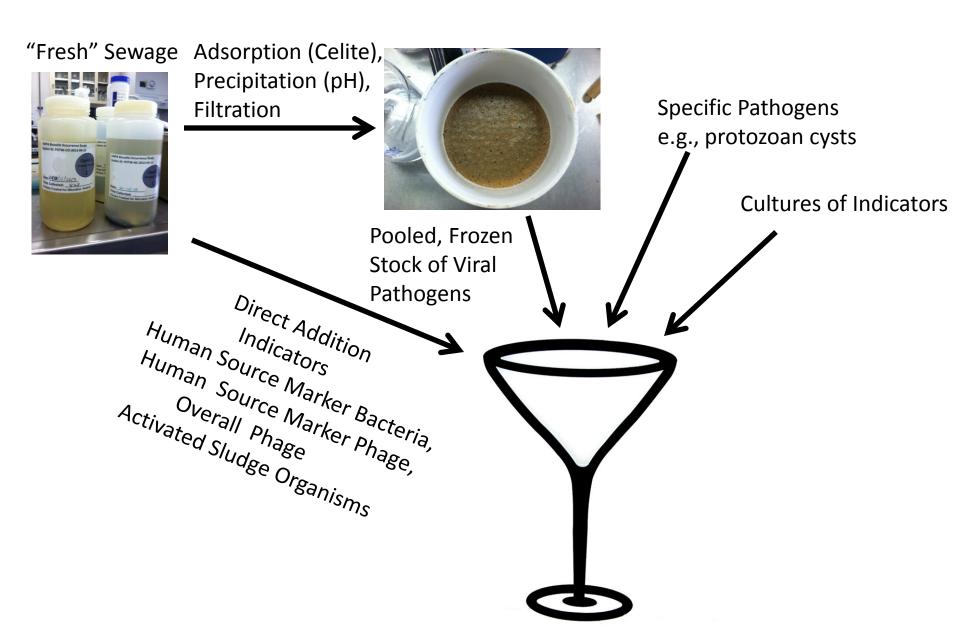
- 248,000 sequences from 12 GW samples
- Range of sequences per sample
 - 13,173 to 37,592
- Range of genera detected per sample
 - 53 to 122
- 97% of the GW sequences were classified as
 - Proteobacteria (209,000 sequences)
 - Bacteroidetes (26,000)
 - Firmicutes (6,300)

196 genera

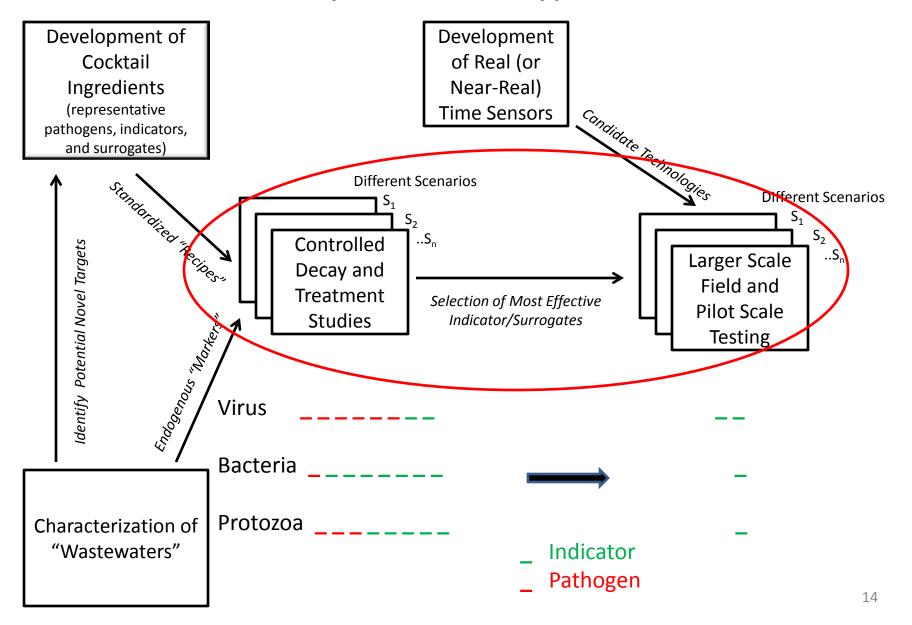
Table 2. List of Genera Classified from Graywater								
Abiotrophia	Cloacibacterium	Lactobacillus	Pseudonocardiaceae					
Achromobacter	chromobacter Clostridium		Pseudorhodoferax					
Acidaminobacter	Coenonia	Leclercia	Pseudoxanthomonas					
Acidovorax	Comamonas	Legionella	Quatrionicoccus					
Acinetobacter	Corynebacterium	Leucobacter	Ralstonia					
Actinomyces	Cupriavidus	Levilinea	Raoultella					
Aeromonas	Curvibacter	Luteibacter	Rhizobium					
Alicycliphilus	Daeguia	Lysobacter	Rhodococcus					
Alkanindiges	Dechloromonas	Magnetospirillum	Rhodocyclus					
Alteromonadales	Deefgea	Massilia	Rhodoplanes					
Amaricoccus	Defluviicoccus	Methylobacterium	Riemerella					
Aminobacterium	Deinococcus	Methylocella	Roseomonas					
Aminomonas	Delftia	Methylocystis	Rothia					
Anaerobacter	Derxia	Methyloversatilis	Rugamonas					
Anaerococcus	Desulfobulbus	Microbacterium	Schlegelella					
Anaerofilum	Desulfocurvus	Microvirga	Serpens					
Anaerovorax	Desulforegula	Microvirgula	Simplicispira					
Ancylobacter	Desulfovibrio	Mitsuaria	Sinobacteraceae					
Aquabacterium	Diaphorobacter	Mycobacterium	Sinorhizobium					
Aquaspirillum	Dokdonella	Nakamurella	Solimonas					
Aquincola	Duganella	Neisseria	Soonwooa					
Aurantimonas	Dyella	Nocardioidaceae	Spartobacteria					
Azomonas	Dysgonomonas	Novispirillum	Sphingobacterium					
Azonexus	Elizabethkingia	Novosphingobium	Sphingobium					
Azorhizobium	Elusimicrobium	Nubsella	Sphingomonas					
Azospira	Enhydrobacter	Oceanospirillales	Sphingopyxis					
Azospirillum	Enterobacter	Olsenella	Sphingosinicella					
Azotobacter	Enterococcus	Opitutus	Spirochaeta					
Azovibrio	Epilithonimonas	Oribacterium	Staphylococcus					
Bacteriovorax	Eubacterium	Ottowia	Stenotrophomonas					
Bacteroides	Ferribacterium	Paludibacterium	Streptobacillus					
Bdellovibrio	Filimonas	Parabacteroides	Streptococcus					
Beijerinckia	Finegoldia	Paracoccus	Sulfuricurvum					
Bellilinea	Flavobacterium	Parvimonas	Sulfurospirillum					
Bilophila	Formivibrio	Pedobacter	Telmatospirillum					
Blastomonas	Fusibacter	Pelomonas	Tessaracoccus					
Bosea	Fusobacterium	Peptoniphilus	Thermomonas					
Brachymonas	Gemella	Peptostreptococcus	Tolumonas					
Bradyrhizobium	Geobacter	Perlucidibaca	Trabulsiella					
Brevundimonas	Geothrix	Phenylobacterium	Treponema					
Brooklawnia	Granulicatella	Phyllobacteriaceae	Uliginosibacterium					
Burkholderia	Haemophilus	Planctomycetaceae	Uruburuella					
Butyrivibrio	Heliothrix	Pleomorphomonas	Variovorax					
Capnocytophaga	Herbaspirillum	Porphyromonas	Victivallis					
Caulobacter	Holophaga	Prevotella	Vogesella					
Chitinimonas	Inquilinus	Prolixibacter	Xanthobacter					
Chryseobacterium	Janthinobacterium	Propionibacterium	Yokenella					
Citrobacter	Klebsiella	Propionivibrio	Zobellella					
Cloacibacillus	Kluyvera	Pseudomonas	Zoogloea					

Overall Research Plan for Developing Tools for Assessing Efficacy of Water Reuse Approaches Development of Development Cocktail of Real (or Ingredients Near-Real) Candidate Technologies (representative Time Sensors pathogens, indicators, and surrogates) Standardized Recipes, **Different Scenarios Different Scenarios** S_1 S_2 S_2 $..S_n$ Controlled Identify Potential Novel Targets $..S_n$ Larger Scale Decay and Field and **Treatment** Selection of Most Effective Endogenous Markers" Pilot Scale **Studies** Indicator/Surrogates Testing Characterization of "Wastewaters" 12

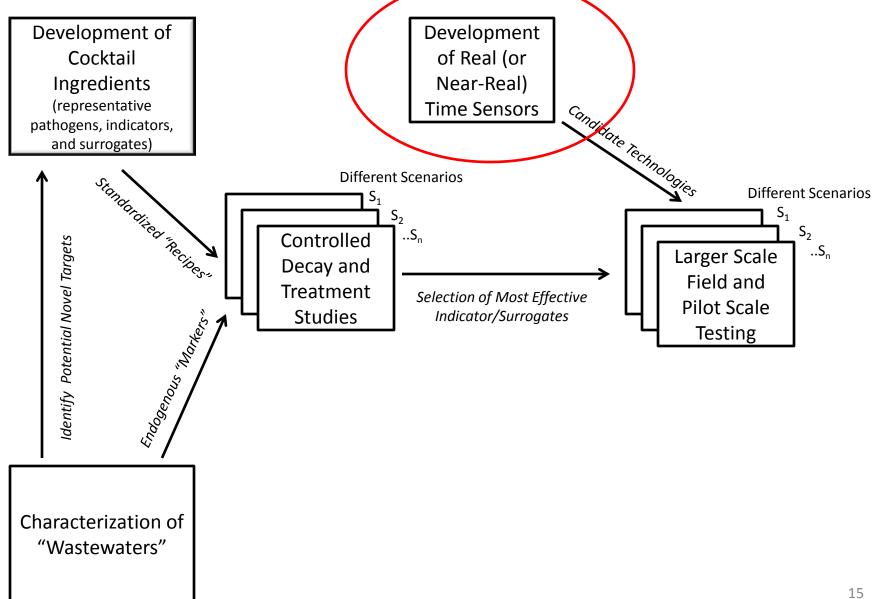
What's the Best Way to Make a Microbial Cocktail?



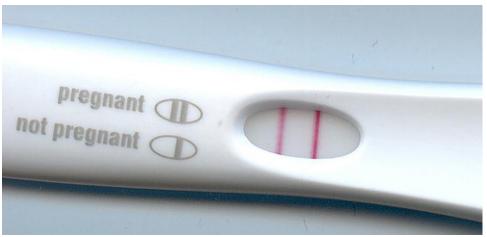
Overall Research Plan for Developing Tools for Assessing Efficacy of Water Reuse Approaches



Overall Research Plan for Developing Tools for Assessing Efficacy of Water Reuse Approaches

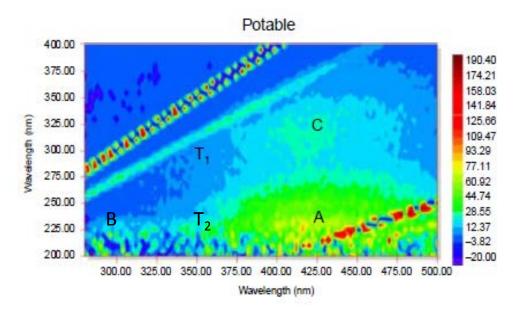


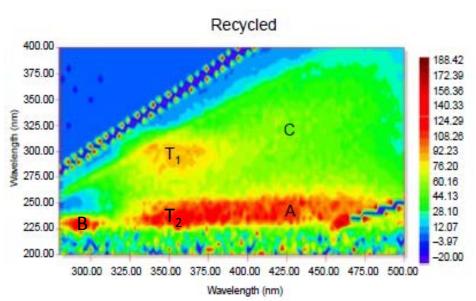




Verifying Quality in Sequential Batches

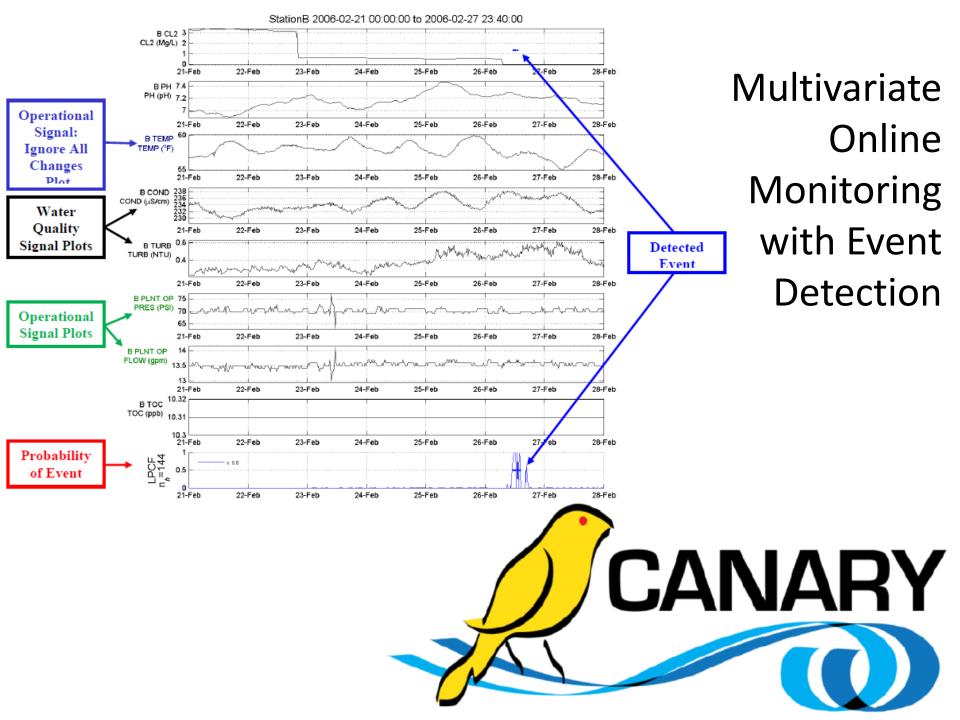
- Rapid "real-time" detection
 - Enzymatic
 - Signal amplification





Peak definitions (Henderson, 2009)

- A: Humic-like
- B: Tryosine-like
- C: Humic-like
- T1: Tryptophan-like
- T2: Tryptophan-like
- Extracellular proteins are mainly excreted by microorganisms. Tryptophan fluorescence is the dominant part of the protein fluorescence, which has a fluorescence maximum at Peak T1 and T2 (Ni, 2009)
- Peak T2 fluorescence correlates with HB, TC, E. Coli (R² values of .81, .78, .72, respectively) from diluted river water and sewage works final effluent (Cumberland, 2012)



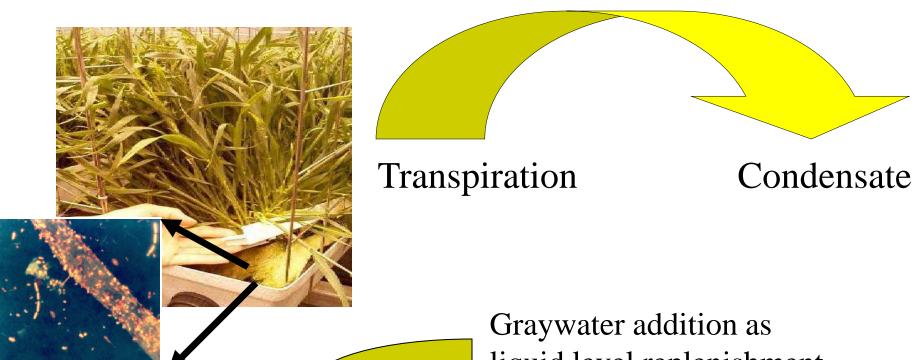
The Need for Eco-Effective Designs for Long Term Human Missions in Space



"If we knew how to live on Mars, we'd know how to reduce our footprint on Earth. Space colonization is the Rosetta stone for earthly sustainability because it's entirely about living in the absence of ecosystem services. The Moon, Mars and the asteroids are a great experimental laboratory that we're ignoring at our own peril."

Karl Schroeder

Direct Graywater Processing Scheme

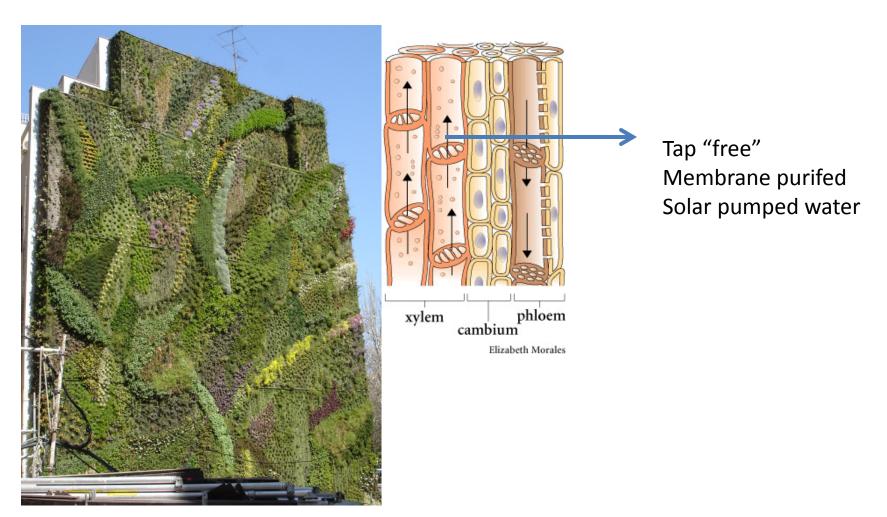


Surfactant Degradation in the Rhizosphere

liquid level replenishment

Maintenance of water quality

Water Research (2004) 38:1952 Water Research (2000) 34:3075



Linking biophillia and low environmental impact design to a new paradigm of sustainable development, referred to as 'restorative environmental design'

Stephen Kellert